Identification of Sex-specific Transcripts of the *doublesex* Gene in the Sawfly, *Athalia rosae ruficornis* (Hymenoptera)^{*1,*2}

Megumi SUMITANI¹⁾, Kazuki SEKINÉ²⁾, Hideki SEZUTSU¹⁾ and Masatsugu HATAKEYAMA²⁾

¹⁾ Genetically Modified Organism Research Center, National Institute of Agrobiological Sciences, Owashi, Tsukuba, Ibaraki 305–8634, Japan

²⁾ Division of Insect Sciences, National Institute of Agrobiological Sciences, Owashi, Tsukuba, Ibaraki 305–8634, Japan E-mail: sumikasashima@affrc.go.jp (MS)

Homologs of the doublesex (dsx) gene of Drosophila melanogaster are highly conserved elements that regulate genetic sex determination in a wide range of organisms (Kopp, 2012). The dsx pre-mRNA is alternatively spliced by transacting proteins, resulting in sex-specific variants. Distinctive dsx isoforms in males and females are known to induce somatic sexual differentiation in a variety of insects (Oliveira et al., 2009). However, functional assays of dsx gene are restricted to Diptera (Baker and Wolfner, 1988; Raymond et al., 1998) and Lepidoptera (Suzuki et al., 2005). Hymenoptera is a unique order in which arrhenotokous parthenogenesis (development of haploid males from unfertilized eggs) is the general mode of reproduction. The sawfly, Athalia rosae, is a new model species in this order that has been used to conduct studies in genetics and developmental biology with tools such as germline transformation by the piggyBac transposonderived vector and gene knockdown by RNAi (Sumitani et al., 2003; Yoshiyama et al., 2013). In addition, its genomic sequence is publicly available (http://www.ncbi.nlm.nih.gov/nuccore/ AOFN00000000.1/). To investigate the regulatory mechanisms of sex determination in A. rosae, we identified its dsx homolog (Ardsx) using whole transcriptome shotgun sequencing (RNA-Seq) (Hatakeyama et al., 2017). In this report, we describe the sex-specific transcripts of A. rosae.

The complete *Ardsx* coding sequence was confirmed using reverse transcription-polymerase chain reaction (RT-PCR) with cDNA constructed from adult females and males as templates. The *Ardsx* gene corresponded to a region spanning approximately 12 kb of the genome. The deduced amino acid sequence showed the typical Dsx structure, with a *doublesex* and *mab-3* (DM) DNA-binding domain (which is characteristic of the zinc-finger motif in the N-terminal region) and a Dsx dimerization domain in the C-terminal region. In males, exons 1, 2, and 3 joined to form a transcript encoding 234 amino acids. In females, specific splicing of 119 nucleotides within exon 3 resulted in the female-specific variant. The resultant female-specific transcript encoded 338 amino acids and had a longer C-terminal region than the male variant. In addition, we identified the *transformer 2 (tra2)* homolog, which is known to regulate sex-specific alternative splicing of *dsx* premRNA in *D. melanogaster* (Ryner and Baker 1991). In contrast, in silkworm (*Bombyx mori*), the *tra2* homolog is not required for sex-specific splicing of *B. mori dsx* pre-mRNA (Suzuki *et al.*, 2012). It would be interesting to determine whether *tra2* is involved in *dsx* splicing in *A. rosae*.

References

- Baker, B. S. and M. F. Wolfner (1988) A molecular analysis of *doublesex*, a bifunctional gene that controls both male and female sexual differentiation in *Drosophila malanogaster*. *Genes and Development*, 2, 477–489.
- Hatakeyama, M., M. Sumitani and K. Sekiné (2017) Identification of gene transcripts using whole transcriptome shotgun sequencing (RNA-Seq) in the sawfly *Athalia rosae ruficornis* (Hymenoptera). *Proceedings of the Arthropodan Embryological Society of Japan*, 48, 53–54.
- Kopp, A. (2012) Dmrt genes in the development and evolution of sexual dimorphism. Trends in Genetics, 28, 175–184.
- Oliveira, D. C., J. H. Werren, E. C. Verhulst, J. D. Girbel, A. Kamping, L. M. Beukeboom and L. van de Zande (2009) Identification and characterization of the *doublesex* gene on *Nasonia*. *Insect Molecular Biology*, 18, 315–324.
- Raymond C. S., C. E. Shamu, M. M. Shen, K. J. Seifert, B. Hirsch, J. Hodgkin and D. Zarkower (1998) Evidence for evolutionary conservation of sex-determining genes. *Nature*, **391**, 691–695.
- Ryner, L. C. and B. S. Baker (1991). Regulation of *doublesex* premRNA processing occurs by 3'-splice site activation. *Genes and Development*, 5, 2071–2085.
- Sumitani, M., D. S. Yamamoto, K. Oishi, J. M. Lee and M. Hatakeyama. (2003) Germline transformation of the sawfly, *Athalia rosae* (Hymenoptera: Symphyta), mediated by a *piggyBac*-derived vector. *Insect Biochemistry and Molecular Biology*, **33**, 449–458.

^{*1} Abstract of paper read at the 49th Annual Meeting of the Arthropodan Embryological Society of Japan, June 7–8, 2013 (Tsukuba-san, Ibaraki).

^{*2} This article, which was accepted in 2013 and should have been published in 2014, was printed in 2017 being much delayed due to various circumstances.

- Suzuki, M. G., S. Funaguma, T. Kanda, T. Tamura and T. Shimada (2005) Role of the male BmDSX protein in the sexual differentiation of *Bombyx mori*. *Evolution and Development*, 7, 58– 68.
- Suzuki, M. G., K. Suzuki, F. Aoki and M. Ajimura (2012) Effect of RNAi-mediated knockdown of the *Bombyx mori transformer-2*

gene on the sex-specific splicing of *Bmdsx pre-mRNA*. The International Journal of Developmental Biology, **56**, 693–699.

Yoshiyama, N., K. Tojo and M. Hatakeyama (2013) A survey of the effectiveness of non-cell autonomous RNAi throughout development in the sawfly, *Athalia rosae* (Hymenoptera). *Journal* of Insect Physiology, **59**, 400–407.